

Hereditary Multiple Exostoses: A Current Understanding of Clinical and Genetic Advances

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Background

Osteochondroma is the most common bone tumor seen in children [6,22,59]. This cartilage-capped exostosis is found primarily at the juxta-epiphyseal region of the most rapidly growing ends of long bones [59,78]. The true prevalence is not known since many patients with asymptomatic lesions are never diagnosed. A unique subset of patients, however, suffers from hereditary multiple exostosis (HME), an autosomal-dominant disorder manifested by multiple osteochondromas and frequently associated with characteristic progressive skeletal deformities. Recent advances in understanding the molecular and genetic basis of this condition not only offer hope for patients and families with HME, but also offer clues to the underlying basis for the formation of the human musculoskeletal system.

Historically, John Hunter was perhaps the first to comment on the condition now known as HME. In 1786, he described a patient with multiple exostoses in his *Lectures on the principles of surgery* [37]. In 1814, Boyer published the first description of a family with HME, and this was followed by Guy's description of a second family in 1825 [9,32]. Most of the clinical aspects of the disease had been described by the late 1800's⁵. HME was introduced into the American literature in 1915 by Ehrenfried. In 1943, Jaffe made a significant contribution by further elucidating the pathology of HME and helping to differentiate the disorder from Ollier's disease [24,38]. As with HME, patients with Ollier's disease have multiple, benign cartilaginous lesions of bone, but the lesions of Ollier's disease are enchondromas, located within the tubular bones.

The name "multiple exostoses" was given to the condition by Virchow in 1876 [92]. A number of synonyms have been used for this disorder including osteochondromatosis, multiple hereditary osteochondromata, multiple congenital osteochondromata, diaphyseal aclasis, chondral osteogenic dysplasia of direction, chondral osteoma, deforming chondrosyplasia, dyschondroplasia, exostosing disease, exos-

totic dysplasia, hereditary deforming chondrodysplasia, multiple osteomatosis, and osteogenic disease [24,35,59]. A related entity known as dysplasia epiphysealis hemimelica, or Trevor's disease, is a rare disorder in which osteochondromas arise from an epiphysis [88].

HME is most frequently described in Caucasians and affects 0.9 to 2 individuals per 100,000; higher prevalences of the condition have been identified in isolated communities such as the Chamorros of Guam or the Ojibway Indian community of Pauingassi in Manitoba, Canada [6,35,42,56,69,85,94]. These populations have a prevalence of 100 and 1310 per 100,000, respectively [6,42]. Although previously thought to have a male predominance [13,38,78], HME now appears to affect both sexes similarly [69,97].

Clinical Presentation

Patients with HME have multiple cartilage-capped exostoses that may be sessile or pedunculated. Although most commonly located at the periphery of the most rapidly growing ends of long bones, the lesions are also frequently found in the vertebral borders of the scapulae, ribs, and iliac crests [79]. Osteochondromas may occur in the tarsal and carpal bones, however they are often less apparent [76] (Fig. 1). There is only one reported case of an exostosis in the skull; there are no reported cases of lesions arising from the facial bones [35,38].

Exostoses are initially recognized and diagnosed in the first decade of life in over 80% of individuals with HME and are most commonly first discovered on the tibia or scapula as these are often the most conspicuous locations [79]. HME is occasionally diagnosed at birth, but such an early diagnosis is usually the result of a specific search—often in the context of a family history of the disorder. Patients with HME vary considerably as to the size and number of lesions. Some individuals have smaller and fewer lesions that may never become symptomatic. The lesions tend to enlarge while the physes are open proportionate to the overall growth of the patient, and the growth of the osteochondromas usually ceases at skeletal maturity. Lesions have been infrequently reported to spontaneously regress during the course of childhood and puberty [13,21]. Recurrence of an exostosis after surgical excision, although rare, has been observed and may be attributed to incomplete removal of lesions contiguous with the physis in growing

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children or incomplete removal of the cartilaginous cap [35].

Clinical Manifestations

While exostoses are histologically and clinically benign lesions, they can result in a variety of problems. Pain, often from soft tissue trauma over exostoses, and cosmetic concerns are frequent complaints in patients with HME. Additionally, bursa formation and resulting bursitis may occur as a result of the exostoses. The most common deformities seen in HME include short stature, limb-length discrepancies, valgus deformities of the knee and ankle, asymmetry of the pectoral and pelvic girdles, bowing of the radius with ulnar deviation of the wrist, and subluxation of the radial head [69,72,79,97]. Relative shortening of the metatarsals, metacarpals, and phalangeas as well as scoliosis, coxa valga, and acetabular dysplasia have been described less commonly [18,28,39,71]. Associated soft tissue problems include tendon, nerve or vascular impingement, entrapment or injury. Spinal cord compression is also a rare, but well documented, complications of HME [26,63,72,73]. Solomon reported both urinary and intestinal obstruction as other uncommon soft tissue complications [79]. Dysphagia secondary to a ventral cervical exostosis and spontaneous hemothorax as a result of rib exostoses have been described

[2,17,23,89]. There have also been reports of exostoses interfering with normal pregnancy and leading to a higher rate of Cesarean sections [46,97].

Individuals with HME are frequently of short stature, with most having heights 0.5 to 1.0 SD below the mean [71,75]. Affected adult males and females have been observed to have heights below the fifth percentile in 36.8% and 44.2% of cases, respectively [97]. Sitting height is generally less abnormal than total height, indicating that the limbs are involved disproportionately as compared to the spine [71].

Limb-length discrepancy is also common. A clinically significant inequality of 2 cm or greater has been reported with a prevalence ranging from 10%–50% [69,71]. Shortening can occur in the femur and/or the tibia; the femur is affected approximately twice as commonly as the tibia [71]. Surgical treatment with appropriately timed epiphysiodesis has been satisfactorily employed in growing patients.

In addition to limb-length discrepancies, a number of lower extremity deformities have been documented. Since the disorder involves the most rapidly growing ends of the long bones, the distal femur is among the most commonly involved sites and 70%–98% of patients with HME have lesions (Fig. 2) [69,71,79]. Coxa valga has been reported in up to 25% [71]; lesions of the proximal femur have been reported in 30%–90% of patients with HME [69,79]. Femoral anteversion and valgus have been associated with exostoses located in proximity to the lesser trochanter [94]. Lesions of the proximal femur can also result impaired hip flexion. There have been at least three reported cases of acetabular dysplasia with subluxation of the hip in patients with HME [28,86]. This results from exostoses located within or about the acetabulum that may interfere with normal articulation.

Valgus knee deformities are found in 8%–33% of patients with HME [56,69,71]. Although distal femoral involvement is common, the majority of cases of angular limb deformities are due mostly to lesions of the proximal tibia and fibula which occur in 70%–98% and 30%–97% of cases, respectively [69,71,79]. The fibula has been found by Nawata et al. to be shortened disproportionately as compared to the tibia, and this is likely responsible for the consistent valgus direction of the deformity [56]. Seven of twenty patients with this valgus deformity in the series by Shapiro et al. required corrective osteotomy [71]. It should be noted that this procedure is associated with appreciable risk due to the proximity of neurovascular structures.

Valgus deformity of the ankle is also common in patients with HME and is observed in 45%–54% of patients in most series [39,71,75]. This valgus deformity can be attributed to multiple factors including shortening of the fibula relative to the tibia (Fig. 3). A resulting obliquity of the distal tibial epiphysis and medial subluxation of the talus can also be associated with this deformity, while developmental obliquity of the superior talar articular surface may provide partial compensation [71]. In more advanced cases, excision of exostoses, alone, does not correct the ankle deformity, although, it may improve preoperative symptoms and cosme-



Fig. 1. CT image through the hindfoot showing a tarsal osteochondroma extending from the infralateral border of the talus.

sis [74]. Early medial hemiepiphyseal stapling of the tibia in conjunction with exostosis excision can correct a valgus deformity at the ankle of 15° or greater associated with limited shortening of the fibula [71,74]. Fibular lengthening has been used effectively for severe valgus deformity with more significant fibular shortening (i.e., when the distal fibular physis is located proximal to the distal tibial physis) [74]. Supramalleolar osteotomy of the tibia has also been used effectively to treat severe valgus ankle deformity [71]. Growth of exostoses can also result in tibiofibular diastasis, which can be treated with early excision of the lesions [80].

Osteochondromas of the upper extremities frequently cause forearm deformities. The prevalence of such deformities has been reported to be as high as 40%–60% [38,69,71,79,100]. Disproportionate ulnar shortening with relative radial overgrowth has been frequently described and may result in radial bowing. Subluxation or dislocation of the radial head is a well-described sequelae in the context of these deformities and was seen in 8 of 37 elbows examined by Shapiro et al. [71] (Fig. 4). Dislocation of radial head has been associated with a loss of pronation, greater ulnar variance, and functional impairment [81]. Disruption of the radioulnar joint, ulnar deviation, and ulnar translocation of the carpus are often associated with HME [27,79]. This complex of deformities, while similar to Madelung's deformity, does not manifest itself in the characteristic rela-

tive elongation or dorsal subluxation of the distal ulna as seen in Madelung's deformity [65,71].

In 1891, Bessel-Hagen was first to discuss deformities of the forearm in HME and proposed that the irregular eccentric growth of osteochondromas accounted for the loss in longitudinal growth of the bone [5]. This hypothesis was supported by Jaffe, and later by Porter et al., who found that the length of forearm bones inversely correlates with the size of the exostoses [38,62]. Thus, the larger the exostoses and the greater the number of exostoses, the shorter the involved bone. Moreover, lesions with sessile rather than pedunculated morphology have been associated with more significant shortening and deformity [16]. Thus, the skeletal growth disturbance observed in HME is a local effect of benign growth [62].

Accordingly, the disproportionate shortening of the ulna can be generally attributed to two causes: since the distal ulnar physis is responsible for greater longitudinal growth relative to that of the distal radius (85% versus 75%), equal involvement results in more substantial ulnar shortening. Additionally, bones with smaller cross-sectional diameter tend to be shortened more considerably when affected by HME, and so equal involvement of the two bones preferentially affects the ulna which has a diameter of only one-fourth that of the radius. Consequently, radial bowing had been theorized to result from a tethering effect due to the

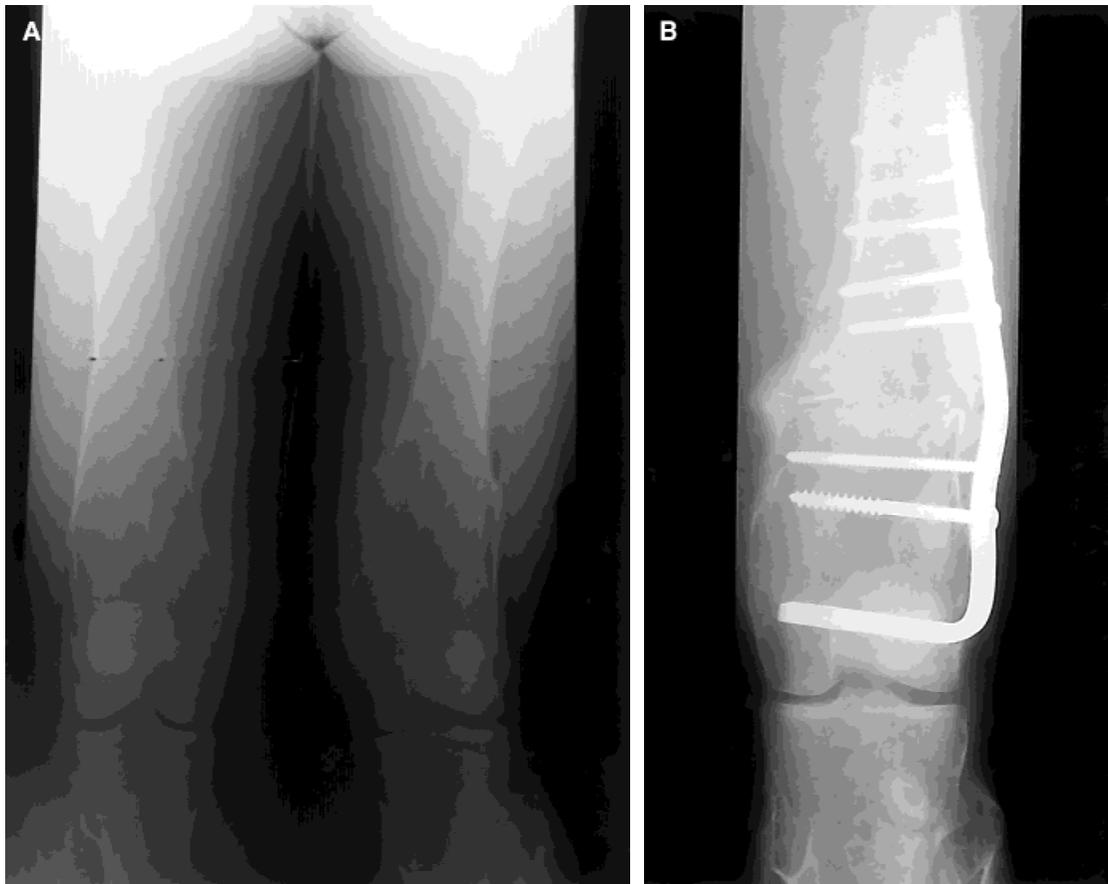


Fig. 2. (A) Standing A/P radiograph of the lower extremities showing left genu valgum. (B) Standing A/P radiograph demonstrating correction of the femoral deformity with a lateral opening-wedge osteotomy and internal fixation.



Fig. 3. A/P radiograph of an ankle demonstrating osteochondromas of the distal tibia and fibula with relative fibular shortening resulting in valgus angular deformity.

relative shortening of the ulna [75,79]. Burgess and Cates, however, disputed this theory with their finding that radial bowing was uncorrelated with measured ulnar shortening in their series of 35 patients, though their study did find a strong correlation between ulnar shortening in excess of 8% and dislocation of the radial head [11].

The degree of forearm involvement in patients with HME has been shown to be strongly associated with the general severity of the disease. Taniguchi classified his patients into three groups: (1) those with no involvement of the distal forearm, (2) those with involvement of the distal radius or ulna without shortening of either bone, and (3) those with involvement of the distal radius or ulna with shortening of either bone. He found that increasing forearm involvement was associated with an earlier age of diagnosis of HME, a greater number of generalized exostoses, shorter stature, a greater number of exostoses affecting the knee, and increased valgus deformity of the ankle. Not surprisingly, all patients with dislocations of the radial head in his series were in the most severely affected group, with shortening in addition to distal exostoses (i.e., group 3) [87].

Many of the deformities of the forearm in patients with HME are amenable to surgical treatment. Indications for surgical treatment include painful lesions, an increasing radial articular angle, progressive ulnar shortening, excessive carpal slip, loss of pronation, and increased radial bowing with subluxation or dislocation of the radial head [99]. In a study of 25 patients who underwent surgery for correction of forearm deformities, Fogel et al. determined that, while

early osteochondroma excision alone may decrease or halt progression of forearm deformity, it did not consistently provide full correction. They found that ulnar translocation of the carpals on the distal radius can be corrected by ulnar lengthening, but persistent relative ulnar shortening is likely to recur (Fig. 5). For patients with increased radiocarpal angulation or carpal subluxation, they concluded that osteochondroma excision in conjunction with distal radial osteotomy or hemiepiphyseal stapling resulted in improved function and cosmesis [29]. Wood et al. noted that such surgeries of the distal forearm result in only modest improvement of function, but they felt, significant improvement in cosmesis [100].

Complete dislocation of the radial head is a serious progression of forearm deformity and can result in pain, instability, and decreased motion at the elbow. Surgical intervention should be considered to prevent this from occurring. When symptomatic, this can be treated in older patients with resection of the radial head [53,71]. Surgical relocation of the radial head, however, has not consistently proven to be successful [100].

Hand involvement in HME has been reported in 30%–79% of patients [69,79]. Fogel et al. observed metacarpal involvement and phalangeal involvement in 69% and 68%,



Fig. 4. A/P radiograph of the forearm showing ulnar shortening and radial head dislocation.



Fig. 5. (A) P/A radiograph of distal forearms showing characteristic radial bow and ulnar shortening of the left wrist. (B) Early postoperative P/A radiographs of the left wrist following corrective osteotomy and pinning. (C) One-year follow-up P/A radiographs of the same wrist showing radial correction with residual ulnar shortening.

respectively, in their series of 51 patients [29]. In their series of 63 patients, Cates and Burgess found that patients with HME fall into two groups: those with no hand involvement and those with substantial hand involvement averaging 11.6 lesions per hand [18]. They documented involvement of the ulnar metacarpals and proximal phalanges most commonly with the thumb and distal phalanges being affected less frequently. While exostoses of the hand resulted in shortening of the metacarpals and phalanges, brachydactyly was also observed in the absence of exostoses [18]. In most series, the majority of patients were asymptomatic [18,71]. In Cates and Burgess's study, no angular deformities of the

digits were observed, and only 4 of 22 patients with hand involvement required surgery [18].

Both neurologic and vascular problems can arise throughout the extremities as complications of HME. Wicklund et al. reported peripheral nerve compression symptoms in 22.6% of patients in their series of 180 [97]. Peroneal neuropathy associated with exostoses of the proximal fibula in children is a recognized complication [14,47]. At our institution, six children were described with peroneal nerve palsy associated with osteochondromas of the proximal fibula [14]. Ulnar neuropathy secondary to compression by an exostosis of the elbow has also been described [68].

Wicklund et al. reported the general prevalence of vascular compression secondary to exostoses to be 11.3% [97]. In their review of vascular complications stemming from osteochondromas, Vasseur et al. reported 97 cases, of which 71 were sporadic osteochondromas while 26 were associated with HME [91]. Pseudoaneurysm, vascular compression, arterial thrombosis, aneurysm, and venous thrombosis were the most commonly reported, while claudication, acute ischemia, and phlebitis were found to be the most commonly associated clinical presentations. In Vasseur et al.'s series, 83% of vascular problems were located in the lower extremity, and the popliteal artery was the most frequently injured artery [91]. Appropriate, and usually urgent, surgical treatment of these patients is required in this setting.

Malignant transformation of a benign osteochondroma to a chondrosarcoma or other sarcoma is another complication of HME. Fortunately, most chondrosarcomas in this setting are low grade and can be treated with wide excision. Patients with such lesions usually present with a painful mass. Rarely, nerve compression can be the presenting complaint [58]. Ochsner published a report of 59 patients with HME who had malignant transformation. The mean age at diagnosis of malignancy was 31 years of age with malignant degeneration seldom occurring in the first decade or after the fifth decade of life [57]. The reported incidence of malignant degeneration is highly variable, ranging from 0.5%–25% [30,77,94]. This disparity can be attributed not only to a possible selection bias inherent for a tertiary referral center, but also to the inability to detect all HME patients without malignant degeneration, thus making it difficult to determine the true denominator [16]. More recent studies estimate the rate of secondary malignancy to be 5% or less [6,31,46,69,94]. The risk of malignant transformation may vary among families reflecting genetic heterogeneity predisposing to malignant degeneration [69]. Because of this risk, patients with HME should be followed carefully to detect early sarcomatous transformation. Growth of a lesion after skeletal maturity should raise a suspicion of malignancy. Additionally, the presence in an adult of an osteochondroma with a cartilaginous cap greater than 2 cm has been associated with an increased chance of malignancy [22].

Genetic Basis of Disease

One of the early studies that looked at the hereditary characteristics of patients with HME was done by Stocks and Barrington in 1925 [84]. Since that time, it has been determined that HME is an autosomal dominant disorder with near complete penetrance [35,78,97]. HME is a genetically heterogeneous disorder and has been associated with mutations in at least three different genes, termed EXT genes. At least two of these genes are thought to function as tumor suppressor genes. The three described EXT loci have been recently mapped: EXT1 on chromosome 8q23-q24 [20], EXT2 on 11p11-p12 [102,104], and EXT3 on chromosome 19p [45]. According to linkage analysis, the EXT1 and EXT2 loci appear to be altered in the majority of fami-

lies while, EXT3, which has not been fully isolated and characterized, is probably less frequently affected [105]. Epidemiologic analysis of linkage and mutation data indicate that mutations of EXT1 and EXT2 are likely to be responsible respectively for one half and one third of multiple hereditary exostoses cases [60,61,67,96,106].

EXT1 and EXT2 function as tumor suppressor genes encoding homologous glycoproteins of similar size (746 and 718 amino acids, respectively) and structure which are expressed ubiquitously throughout the musculoskeletal system [1,82,109]. Both glycoproteins are glycosyltransferases that function in the biosynthesis of heparan sulfate [52]. They are located in the membrane of the endoplasmic reticulum and have a role in modifying and enhancing the synthesis and expression of heparan sulfate, a complex polysaccharide that has been implicated in a variety of cellular processes including cell adhesion, growth factor signaling, and cell proliferation [15,101].

Wuyts and Van Hul proposed a model for the development of exostoses based upon a mutation in the EXT gene [105]. They note that the function of the EXT gene may be better understood by studying the *tout-velu* gene, the *Drosophila* homologue of EXT1. The *tout-velu* gene has been implicated in the normal diffusion of hedgehog (*hh*), a signaling protein [4]. Among the mammalian homologues of hedgehog is Indian hedgehog (*Ihh*), a regulator of cartilage differentiation. Indian Hedgehog is expressed by chondrocytes and then diffuses into the perichondrium. There, it exerts its influence by inhibiting further differentiation of additional chondrocytes [93]. Wuyts and Van Hul offer a mechanism of exostosis formation in which a mutation in the EXT gene disrupts Indian hedgehog diffusion, in turn, inhibiting the negative feedback loop present in chondrocyte differentiation and resulting in abnormal skeletal development [105]. Thus, a mutation in the EXT gene may disrupt normal cartilage growth resulting in the formation of an osteochondroma.

Three other homologous genes, termed EXT-like genes, have been identified: EXTL1 on chromosome 1p36 [99], EXTL2 on 1p11-p12 [107], and EXTL3 on 8p12-p22 [90]. Unlike EXT1 and EXT2, the EXTL proteins are more variable in size. They do however, share characteristic features with the EXT gene family such as conserved biologically active sequences. Most notably, EXT2 has been demonstrated to be a transferase involved in the biosynthesis of heparin sulfate and likely encodes the crucial enzyme, which initiates heparan sulfate synthesis [41]. While EXTL2 has been implicated in the same pathway as EXT1 and EXT2, its role in the formation of exostoses remains to be proven [105].

HME can also be associated with certain other genetic syndromes. While the EXT phenotype resulting from small insertions, deletions, and point mutations is limited to the growth of exostoses, more substantial deletions involving the EXT1 or EXT2 genes in addition to other adjacent genes can result in continuous gene syndromes. Such syndromes are caused by larger deletions which inactivate several genes in the germline. Multiple exostoses are seen in patients with

Langer-Giedion syndrome (LGS), or tricho-rhino phalangeal syndrome type II (TRPII), and DEFECT 11 syndrome. Along with exostoses due to the deletion of the EXT1 gene, patients with TRPII commonly display mental retardation, cone-shaped epiphyses, and atypical facies [43]. This syndrome is caused by deletion of the yet to be mapped TRPI gene which is located proximally to the EXT1 locus [49,50]. DEFECT 11 syndrome is seen in patients with deletions including the entire EXT2 gene on chromosome 11p11-p12. This syndrome is comprised of exostoses in addition to enlarged parietal foramina, craniofacial dysostosis, and mental retardation [3,48,103].

Both EXT1 and EXT2 hereditary multiple exostoses pedigrees exhibit germline mutations in the EXT gene that consist primarily of loss-of-function mutations, often resulting in premature stop codons [1,19,60,83,109]. These mutations result in truncated proteins with decreased biological activity. Further examination shows that in both sporadic and inherited exostoses, chromosomal deletions are present surrounding the EXT1 and EXT2 loci [54]. Additionally, the EXT-like genes are located at sites of tumor suppressor genes in neoplasia; EXTL1 has been localized to 1p36, which is often a site of deletion in tumors, and EXTL3 may be a breast cancer locus [90,99].

As further evidence for a role in tumor suppression, a number of studies have demonstrated loss of heterozygosity (LOH) at the EXT loci in the cartilaginous cap of osteochondromas and tissue from chondrosarcomas [7,8,33,34,66]. Clonal karyotypic abnormalities have also been documented in osteochondromas [10,54]. Taken together, these studies indicate that the cartilaginous portion of the osteochondroma has a clonal or neoplastic origin [8]. Porter and Simpson then contend that the 'osteal' portion of the osteochondroma functions as reactive or supportive stroma since it has been observed that surgical ablation of the cartilage cap alone prevents continued growth of the lesion [62]. There is currently, however, no molecular or immunohistochemical data which supports this observation [8].

As evidence for a genetic progression model of tumor formation, osteochondroma occurrence in HME requires inactivation of both copies of the EXT1 gene in cartilaginous cells [8,9,36]. It remains unclear whether this complete inactivation also occurs in sporadic osteochondromas [8]. The process of malignant transformation to a peripheral chondrosarcoma from a benign precursor may require additional genetic alterations. Additionally, there exists some evidence that carcinogenesis may be associated with deletions found in osteosarcoma and in multiple endocrine neoplasia [61]. Aggressive chondrosarcomas have also been associated with p53 tumor suppressor gene deletions [66]. The genetic changes which accompany malignant transformation require further study before the true mechanism is fully understood.

Pathogenesis

Although the true pathogenesis of HME is not fully understood, many theories have been proposed. Isolation of

cartilaginous islets from the diaphyseal surface of growing cartilage had been hypothesized to cause abnormal osteogenesis. Further, physical stress at sites of tendon attachment had also been thought to convert focal accumulations of embryonic connective tissue to hyaline cartilage. Anatomical theories have attributed osteochondroma formation to a defect in the anchoring of germinal cartilage cells to the physis or to a failure of a thin cortical sleeve of bone acting as a structural constraint allowing a spill-over of physal cells onto the metaphysis [40,61,64,79].

Theories of pathogenesis still exist which remain consistent with a clonal etiology. Müller theorized that osteochondromas result from a primary defect in periosteal differentiation in which ectopic collections of cartilage cells arise from the proliferative layer of the metaphyseal periosteum [38,55]. Osteochondromas have also been thought to arise from multipotent mesenchymal cells in the region of the perichondrial groove of Ranvier [12,70], or, according to Langenskiöld, from proliferative interstitial physal chondrocytes that persist in chondrogenesis as they are transformed into the proliferative layer of the metaphyseal periosteum [44].

Conclusions

In order to better understand HME, additional research is required. It still remains unclear as to why EXT expression is so widespread in human tissues, yet only inactivation at specific sites results in defects. Additionally, the role of impaired heparan sulfate expression will need to be further explored so that the pathogenesis of the disorder can be determined. The EXT3 gene remains to be fully characterized along with other genes possibly involved in HME, and functional analysis of these genes, in addition to the EXT1 and EXT2 genes already identified, is an area of current research [19,51,83]. The genotypic-phenotypic relationship in HME is also being actively investigated [16]. While much has already been learned about HME, further elucidation of the genetic basis and pathogenesis holds promise for better prediction of prognosis and treatment, and, perhaps, may provide additional information about the mechanisms and secrets of normal limb development or other musculoskeletal disorders.

References

1. Ahn J, Lüdecke HY, Lindow S, Horton WA, Lee B, Wagner MJ, Horsthemke B, Wells DE. Cloning of the putative tumour suppressor gene for hereditary multiple exostoses (EXT1). *Nat Genet* 11:137-43, 1995.
2. Barros FTE, Oliveira RP, Taricco MA, Gonzalez CH. Hereditary multiple exostoses and cervical ventral protuberance causing dysphagia: A case report. *Spine* 20:1640-2, 1995.
3. Bartsch O, Wuyts W, Van Hul W, Hecht JT, Meinecke P, Hogue D, Werner W, Zabel B, Hinkel GK, Powell CM, et al. Delineation of a contiguous gene syndrome with multiple exostoses, enlarged parietal foramina, craniofacial dysostosis and mental retardation, caused by deletions on the short arm of chromosome 11. *Am J Hum Genet* 58:734-42, 1996.

4. Beilaiche Y, The I, Perrimon N. Tout-velu is a *Drosophila* homologue of the putative tumour suppressor EXT-1 and is needed for Hh diffusion. *Nature* 394:85–8, 1998.
5. Bessel-Hagen F. Deformity of the forearm. *Arch Klin Chir* 41:42–466, 1891.
6. Black B, Dooley J, Pyper A, Reed M. Multiple hereditary exostoses: An epidemiologic study of an isolated community in Manitoba. *Clin Orthop* 287:212–17, 1993.
7. Bovée J, Cleton-Jansen A, Wuyts W, Caethoven G, Taminiau A, Bakker E, Van Hul W, Cornelisse P, Hogendoorn P. EXT mutation analysis and loss of heterozygosity in sporadic and hereditary osteochondromas and secondary chondrosarcomas. *Am J Hum Genet* 65: 689–98, 1999.
8. Bovée J, Hogendoorn P. Re. Review article entitled 'The neoplastic pathogenesis of solitary and multiple osteochondromas'. *J Pathol* 190: 516–17, 2000.
9. Boyer, A. *Traité des maladies chirurgicales. Paris Ve Migneret* 3: 594, 1814.
10. Bridge JA, Nelson M, Orndal C, Bhatia P, Neff JR. Clonal karyotypic abnormalities of the hereditary multiple exostoses chromosomal loci 8q24.1 (EXT1) and 11p11–12 (EXT2) in patients with sporadic and hereditary osteochondromas. *Cancer* 82:1657–63, 1998.
11. Burgess RC, Cates H. Deformities of the forearm in patients who have multiple cartilaginous exostosis. *J Bone and Joint Surg* 75-A: 13–18, 1993.
12. Burkus JK, Ogdan JA. Development of the distal femoral epiphysis: a microscopic morphological investigation of the zone of Ranvier. *J Ped Orthop* 4:661–8, 1984.
13. Canella P, Gardini F, Boriani S. Exostosis: development, evolution and relationship to malignant degeneration. *Italian J Orthop Traumatol* 7:293–8, 1981.
14. Cardelia JM, Dormans JP, Drummond DS, Davidson RS, Duhaime C, Sutton L. Proximal fibular osteochondroma with associated peroneal nerve palsy: A review of six cases. *J Ped Orthop* 15:574–7, 1995.
15. Carey DJ. Syndecans: multifunctional cell-surface co-receptors. *Biochem* 327:1–16, 1997.
16. Carroll KL, Yandow SM, Ward K, Carey JC. Clinical correlation to genetic variations of hereditary multiple exostosis. *J Ped Orthop* 19:785–91, 1999.
17. Castells L, Comas P, Gonzalez A, Vargas V, Guardia J, Gifre L. Case report: haemothorax in hereditary multiple exostosis. *British J Rad* 66:269–70, 1993.
18. Cates HE, Burgess RC. Incidence of brachydactyly and hand exostosis in hereditary multiple exostosis. *J Hand Surg* 16A:127–32, 1991.
19. Clines GA, Ashley JA, Shah S, Lovett M. The structure of the human multiple exostoses 2 gene and characterization of homologs in mouse and *Caenorhabditis elegans*. *Genome Res* 7:359–67, 1997.
20. Cook A, Raskind W, Blanton SH, Pauli RM, Gregg RG, Francoman CA, Puffenberger E, Conrad EU, Schmale G, Schellenberg G, et al. Genetic heterogeneity in families with hereditary multiple exostoses. *Am J Hum Genet* 53:71–9, 1993.
21. Copeland RL, Meehan PL, Morrissy RT. Spontaneous regression of osteochondromas. *J Bone Joint Surg* 67:971–3, 1985.
22. Dahlin DC, Unni KK: *Bone Tumors: General Aspects and Data on 11,087 Cases*. 5th ed. Philadelphia: Lippincott-Raven; p 4, 11–23, 1996.
23. Dendale J, Amram S, Dermakar S, Guilaud R, Lesbros D. A rare complication of multiple exostoses: hemothorax. *Archives de Pediatie* 2:548–50, 1995.
24. Ehrenfried A. Multiple cartilaginous exostoses—hereditary deforming chorodysplasia: A brief report on a little know disease. *JAMA* 64:1642–6, 1915.
25. Eder HG, Oberbauer RW, Ranner G. Cervical cord compression in hereditary multiple exostoses. *J Neurosurgical Sciences* 37:53–6, 1993.
26. Ergun R, Okten AI, Beskonakli E, Akdemir G, Taskin Y. Cervical laminar exostosis in multiple hereditary osteochondromatosis: anterior stabilization and fusion technique for preventing instability. *European Spine J* 6:267–9, 1997.
27. Fairbank HAT. Diaphyseal aclasis. *J Bone Joint Surg* 31-B:105–13, 1949.
28. Felix NA, Mazur JM, Loveless EA. Acetabular dysplasia associated with hereditary multiple exostoses: A case report. *J Bone Joint Surg* 82-B:555–7, 2000.
29. Fogel GR, McElfresh EC, Peterson HA, Wicklund PT. Management of deformities of the forearm in multiple hereditary osteochondromas. *J Bone Joint Surg* 66-A:670–80, 1984.
30. Garrison RC, Unni KK, McLeod RA, Pritchard DJ, Dahlin DC. Chondrosarcoma arising in osteochondroma. *Cancer* 49:1890–7, 1982.
31. Gordon SL, Buchanan JR, Ladda RL. Hereditary multiple exostoses: report of a kindred. *J Med Genet* 18:428–30, 1981.
32. Guy's Hospital Reports Case of Cartilaginous Exostosis. *Lancet* 2:91, 1825.
33. Hecht JT, Hogue D, Strong LC, Hansen MF, Blanton SH, Wagner M. Hereditary multiple exostosis and chondrosarcoma: linkage to chromosome II and loss of heterozygosity for EXT-linked markers on chromosomes II and 8. *Am J Hum Genet* 56:1125–31, 1995.
34. Hecht JT, Hogue D, Wang Y, Blanton S, Wagner M, Strong LC, Raskind W, Hansen MF, Wells D. Hereditary multiple exostoses (EXT): Mutational studies of familial EXT1 cases and EXT-associated malignancies. *Am J Hum Genet* 60:80–6, 1997.
35. Hennekam RCM. Hereditary multiple exostoses. *J Med Genet* 28: 262–6, 1991.
36. Hogue DA, Cole WG, Hecht J. Mutational steps leading to the development of bony exostoses. *Am J Hum Genet* 63:A365, 1998.
37. Hunter J: *The works of John Hunter, F.R.S. Vol 1*. Palmer JF, editor. London: Longman, Rees, Orne, Brown, Green, and Longman; 1835.
38. Jaffe H. Hereditary multiple exostosis. *Arch Pathol Lab Med* 36:335, 1943.
39. Jahss MH, Olives R. The foot and ankle in multiple hereditary exostoses. *Foot Ankle* 1:128, 1980.
40. Keith, A. Studies on the anatomical changes which accompany certain growth-disorders of the human body. *J Anat* 54:101, 1920.
41. Kitagawa H, Shimakawa H, Sugahara K. The tumor suppressor EXT-like gene EXTL2 encodes an alpha 1,4-N-acetylhexosaminyltransferase that transfers N-acetyl-galactosamine and N-acetylglucosamine to the common glycosaminoglycan-protein linkage region—the key enzyme for the chain initiation of heparan sulfate. *J Biol Chem* 274:13933–7, 1999.
42. Krooth RS, Macklin MT, Hilbish TF. Diaphyseal aclasis (multiple exostosis) on Guam. *Am J Hum Genet* 13:340–7, 1961.
43. Langer LO, Krassikoff N, Laxova R, Scheer-Williams M, Lutter LD, Gorlin RJ, Jennings CG. The trichorhino-phalangeal syndrome with exostoses (or Langer-Giedion syndrome): four additional patients without mental retardation and review of the literature. *Am J Med Genet* 19:81–111, 1984.
44. Langenskiöld A. The development of multiple cartilaginous exostoses. *Acta Orthop Scand* 38:259–66, 1967.
45. Le Merrer M, Legeai-Mallet L, Jeannin PM, Horsthemke B, Schinzel A, Plauchu H, Toutain A, Achard F, Munnich A, Maroteaux P. A gene for hereditary multiple exostoses maps to chromosome 19p. *Hum Mol Genet* 3:717–22, 1994.
46. Leone NC, Shupe JL, Gardner EJ, Millar EA, Olson AE, Phillips EC. Hereditary multiple exostosis: A comparative human-equine-epidemiologic study. *J Hered* 78:171–7, 1987.
47. Levin KH, Wilbourn AJ, Jones HR Jr. Childhood peroneal neuropathy from bone tumors. *Pediatr Neurol* 7:308–9, 1991.
48. Ligon AH, Potocki L, Shaffer LG, Stickens G, Evans GA. Gene for multiple exostoses (EXT2) maps to 11(p11.2–12) and is deleted in patients with a contiguous gene deletion. *Am J Med Genet* 75:538–40, 1998.
49. Lüdecke HJ, Schmidt O, Nardmann J, von Holtum D, Meinecke P, Muenke M, Horsthemke B. Genes and chromosomal breakpoints in the Langer-Giedion syndrome region on human chromosome 8. *Hum Genet* 105:619–28, 1999.

50. Lüdecke HJ, Wagner MJ, Nardmann J, La Pillo B, Parrish JE, Willems PJ, Haan EA, Frydmann M, Hamers GJH, Wells DE, Horsthemke B. Molecular dissection of a contiguous gene syndrome: localization of the genes involved in Langer-Giedion syndrome. *Hum Mol Genet* 4:31–6, 1995.
51. McCormick C, Duncan G, Tufaro F. New perspectives on the molecular basis of hereditary bone tumours. *Mol Med Today* 5:481–6, 1999.
52. McCormick C, Leduc Y, Martindale D, Mattison K, Esford LE, Dyer AP, Tufaro F. The putative tumour suppressor EXT1 alters the expression of cell-surface heparan sulfate. *Nat Genet* 19:158–61, 1998.
53. McCornack EB. The surgical management of hereditary multiple exostosis. *Ortho Rev* 5:57–63, 1981.
54. Mertens F, Rydholm A, Kriebelberg A, Willen H, Jonsson K, Heim S, Mitelman F, Mandahl N. Loss of chromosome band 8q24 in sporadic osteochondrogenous exostoses. *Gene Chrom Cancer* 9:8–12, 1994.
55. Müller E. Über hereditäre multiple cartilagine exostosen und exchondrosen. *Beitr Pathol Anat* 57:232, 1914.
56. Nawata K, Teshima R, Minamizaki T, Yamamoto K. Knee deformities in multiple hereditary exostoses: A longitudinal radiographic study. *Clin Orthop* 313:194–9, 1995.
57. Ochsner PE. Zum problem der neoplastischen entartung bei multipen kartilaginearen exostosen. *Z Orthop* 116:369–78, 1978.
58. Paik NJ, Han TR, Lim SJ. Multiple peripheral nerve compressions related to malignantly transformed hereditary multiple exostoses. *Muscle Nerve* 23:1290–4, 2000.
59. Peterson HA. Multiple hereditary osteochondromata. *Clin Orthop* 239:222–30, 1989.
60. Phillippe C, Porter DE, Emerton ME, Simpson AHRW, Monaco AH. Mutation screening of EXT1 and EXT2 genes in patients with hereditary multiple exostoses. *Am J Hum Genet* 61:520–8, 1997.
61. Porter DE, Simpson AHRW. The neoplastic pathogenesis of solitary and multiple osteochondromas. *J Pathol* 188:119–25, 1999.
62. Porter DE, Simpson AHRW. Reply to letter Re. Review article entitled 'The neoplastic pathogenesis of solitary and multiple osteochondromas'. *J Pathol* 190:517, 2000.
63. Quirini GE, Meyer JR, Herman M, Russell EJ. Osteochondroma of the thoracic spine: an unusual cause of spinal cord compression. *Am J Neuroradiology* 17:961–4, 1996.
64. Rang M: *The growth plate and its disorders*. Edinburgh: E&S Livingstone; 1969.
65. Ranwat CS, DeFirore J, Straub LR. Madelung's deformity: an end-result study of surgical treatment. *J Bone Joint Surg* 57-A:772–5, 1975.
66. Raskind WH, Conrad EU III, Chansky H, Matsushita M. Loss of heterozygosity in chondrosarcomas for markers linked to hereditary multiple exostoses loci on chromosomes 8 and 11. *Am J Hum Genet* 56:1132–9, 1995.
67. Raskind WH, Conrad EU III, Matsushita M, Wijsman EM, Wells DE, Chapman N, Sandell LJ, Wagner M, Houck J. Evaluation of locus heterogeneity of EXT1 mutations in 34 families with hereditary multiple exostoses. *Hum Mutat* 11:231–9, 1998.
68. Ruth RM, Groves RJ. Synovial osteochondromatosis of the elbow presenting with ulnar nerve neuropathy. *Am J Orthop* 25:843–4, 1996.
69. Schmale GA, Conrad EU III, Raskind WH. The natural history of hereditary multiple exostoses. *J Bone Joint Surg* 76-A:986–92, 1994.
70. Shapiro F, Holtrop ME, Blimcher MJ. Organization and cellular biology of the perichondrial ossification groove of Ranvier. *J Bone Joint Surg* 59-A:703–23, 1978.
71. Shapiro F, Simon S, Glimcher MJ. Hereditary multiple exostoses. *J Bone Joint Surg* 61-A:815–24, 1979.
72. Shapiro SA, Javid T, Putty T. Osteochondroma with cervical cord compression in hereditary multiple exostoses. *Spine* 15:600–2, 1990.
73. Silber JS, Mathur S, Ecker M. A solitary osteochondroma of the pediatric thoracic spine: a case report of and review of the literature. *Am J Orthop* 29:711–14, 2000.
74. Snearly WN, Peterson HA. Management of ankle deformities in multiple hereditary osteochondromata. *J Ped Orthop* 9:427–32, 1989.
75. Solomon L. Bone growth in diphysal aclasis. *J Bone Joint Surg* 43-B:700–16, 1961.
76. Solomon L. Carpal and tarsal exostosis in hereditary multiple exostoses. *Clin Radiol* 18:412–16, 1967.
77. Solomon L. Chondrosarcoma in hereditary multiple exostosis. *S Afr Med J* 48:671–6, 1974.
78. Solomon L. Hereditary multiple exostosis. *Am J Hum Genet* 16:35–363, 1964.
79. Solomon L. Hereditary multiple exostosis. *J Bone Joint Surg* 45-B: 292–304, 1963.
80. Spatz DK, Guille JT, Kumar SJ. Distal tibiofibular diastasis secondary to osteochondroma in a child. *Clin Orthop* 345:195–7, 1997.
81. Stanton RP, Hansen MO. Function of the upper extremities in hereditary multiple exostoses. *J Bone Joint Surg* 78-A:568–73, 1996.
82. Stickens D, Clines G, Burbee D, Ramos P, Thomas S, Hogue D, Hecht JT, Lovett M, Evans GA. The EXT2 multiple exostoses gene defines a family of putative tumour suppressor genes. *Nat Genet* 14:25–32, 1996.
83. Stickens D, Evans GA. Isolation and characterization of the murine homolog of the human EXT2 multiple exostoses gene. *Biochem Mol Med* 61:16–21, 1997.
84. Stocks P, Barrington A: *Hereditary disorders of bone development: The treasury of human inheritance*. Vol. 3. Cambridge: Cambridge University Press; 1925.
85. Sugiura Y, Sugiura I, Iwata H. Hereditary multiple exostosis: diaphyseal aclasis. *Jinrui Idengaku Zasshi* 21:149–67, 1976.
86. Tachdjian ME: *Pediatric orthopedics*. Vol. 2. Philadelphia: W.B. Saunders Company; 1990. p 1172–90.
87. Taniguchi K. A practical classification system for multiple cartilagenous exostosis in children. *J Ped Orthop* 15:585–91, 1995.
88. Trevor D. Tarso-epiphysal aclasis: A congenital error of epiphysal development. *J Bone Joint Surg* 32-B:204–13, 1950.
89. Uchida K, Kurihara Y, Sekiguchi S, Doi Y, Matsuda K, Miyanaga M, Ikeda Y. Spontaneous haemothorax caused by costal exostosis. *European Resp J* 10:735–6, 1997.
90. Van Hul W, Wuyts W, Hendrickx J, Speleman F, Wauters J, De Boule K, Van Roy N, Bossuyt P, Willems PF. Identification of a third EXT-like gene (EXTL3) belonging to the EXT gene family. *Genomics* 47:230–7, 1998.
91. Vasseur MA, Fabre O. Vascular complications of osteochondromas. *J Vasc Surg* 31:532–8, 2000.
92. Virchow R: Ueber th Entstehung des Enchondroms und seine Beziehungen zur Enchondrosis und Exostosis cartilaginea. *Manats-berichte der Koniglichen Preussischen Akademie der Wissenschaften*, 760, 1876.
93. Vortkamp A, Lee K, Lanske B, Segre GV, Kronenberg HM, Tabin CJ. Regulation of rate of cartilage differentiation by Indian hedgehog and PTH-related protein. *Science* 273:613–22, 1996.
94. Voutsinas S, Wynne-Davies R. The infrequency of malignant disease in diaphyseal aclasis and neurofibromatosis. *J Med Genet* 2:345–9, 1983.
95. Weiner DS, Hoyt WA. The development of the upper end of the femur in multiple hereditary exostosis. *Clin Orthop* 137:187–90, 1978.
96. Wells DE, Hill A, Lin X, Ahn J, Brown N, Wagner MJ. Identification of novel mutations in the human EXT1 tumor suppressor gene. *Hum Genet* 99:612–15, 1997.
97. Wicklund CL, Pauli RM, Johnston D, Hecht JT. Natural history study of hereditary multiple exostoses. *Am J Med Genet* 55:43–6, 1995.
98. Willems PJ. Identification and characterization of a novel member of the EXT gene family, EXTL2. *Eur J Hum Genet* 5:382–9, 1997.
99. Wise CA, Clines GA, Massa H, Trask BJ, Lovett M. Identification and localization of the gene for EXTL, a third member of the multiple exostoses gene family. *Genome Res* 7:10–16, 1997.
100. Wood VE, Sauser D, Mudge D. The treatment of hereditary multiple exostosis of the upper extremity. *J Hand Surg* 10-A:505–13, 1985.
101. Woods A, Couchman JR. Syndecans: synergistic activators of cell adhesion. *Trends Cell Biol* 8:189–92, 1998.

102. Wu Y, Heutink P, de Vries B, Sandkuijl LA, van den Ouweland AMX, Niermeijer MF, Galjaard H, Reuniers E, Willems PF, Halley DJJ. Assignment of a second locus for multiple exostoses to the pericentromeric region of chromosome 11. *Hum Mol Genet* 3:167–71, 1994.
103. Wuyts W, Di Gennaro G, Bianco F, Wauters J, Morocutti C, Pierelli F, Bossuyt P, Van Hul W, Casali C. Molecular and clinical examination of an Italian DEFECT 11 family. *Eur J Hum Genet* 7:579–84, 1999.
104. Wuyts W, Ramlakhan S, Van Hul W, Hecht JT, van den Ouweland AMW, Raskind W, Hofstede FC, Reyniers E, Wells DE, de Vries B, et al. Refinement of the multiple exostoses locus (EXT2) to a 3 cM interval on chromosome 11. *Am J Hum Genet* 57:383–7, 1995.
105. Wuyts W, Van Hul W. Molecular basis of multiple exostoses: Mutations in the EXT1 and EXT2 genes. *Hum Mutat* 15:220–7, 2000.
106. Wuyts W, Van Hul W, De Boule K, Hendrickx J, Bakker E, Vanhoenacker F, Mollica F, Ludecke HJ, Sayli BS, Pazzaglia UE, et al. Mutations in the EXT1 and EXT2 genes in hereditary multiple exostoses. *Am J Hum Genet* 62:346–54, 1998.
107. Wuyts W, Van Hul W, Hedrickx J, Wauters J, Speleman F, De Boule K, Bossuyt P, Van Agtmael T, Willems PJ. Identification and characterization of a novel member of the EXT gene family, EXTL2. *Eur J Hum Genet* 5:282–9, 1997.
108. Wuyts W, Van Hul W, Hendrickx J, Wauters J, Speleman F, De Boule K, Bossuyt P, Van Agtmael T, Kitagawa H, Shimakawa H, Sugahara K. The tumor suppressor EXT-like gene EXTL2 encodes an alpha 1,4-N-acetylhexosaminyltransferase that transfers N-acetyl-galactosamine and N-acetylglucosamine to the common glycosaminoglycan-protein linkage region—the key enzyme for the chain initiation of heparan sulfate. *J Biol Chem* 274:13933–7, 1999.
109. Wuyts W, Van Hul W, Wauters J, Nemtsova M, Reyniers E, Van Hul E, De Boule K, de Vries BBA, Hendrickx J, Herrygers I, et al. Positional cloning of a gene involved in hereditary multiple exostoses. *Mol Genet* 5:1547–57, 1996.